

Recent Advances in High Pressure Processing of Milk and Milk Products - A review

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ABSTRACT

As the global consumers' demand towards minimally processed freshlike foods has been continuously increasing, efforts to develop novel food processing technologies have been intensified. Among non-thermal food processing technologies, high pressure processing (HPP) seems to be more advantageous due to its environmentally friendly nature, cost efficiency, suitability for processing foods in any form and its positive impacts on foods'

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shelf-life as well as providing efficient microbial safety. Microbiological inactivation efficiency of HPP has been well documented but the role of this technology in digestion efficiency of milk compounds is yet to be elucidated in detail. Also, the potential safety hazards and challenges of HPP in foods require more intense studies. This review deals with the recent developments in HPP treatment to milk and milk products.

1. Introduction and Technological Background of High Hydrostatic Pressure

Food consumption paradigms have changed dramatically during the last two decades. Today, consumers are keener on consuming safer, healthier and minimally processed fresh-like foods and this demand has become more pronounced during coronavirus disease-2019 pandemic. The efficiency of thermal applications to foods including thermization, pasteurization, ultra-high pasteurization and ultra-high temperature treatments on providing food safety has been known for many decades. Although thermal technologies are widely employed in food processing worldwide, possible adverse effects of heat treatment on some food components have motivated the food community to develop alternative technologies to heating. Efforts have been intensified to adapt some non-thermal food processing technologies from lab/pilot scale to industrial scale, and some non-thermal food processing technologies are now enjoying market success and consumer acceptance. High hydrostatic pressure (HPP), pulsed electric field (PEF), ultrasound, cold plasma, ozonization, irradiation and ultraviolet light are some examples of non-thermal food processing technologies. These technologies are employed in shorter processing time and at lower temperatures which leads to improvement of the nutritional quality of foods as well as sensory quality. Also, the shelf-lives of non-thermally processed foods are improved without impairing the food safety parameters (Mahalik & Nambiar 2010; Alexandre et al. 2012).

HPP has gained popularity in food processing faster than the other non-thermal technologies. About 75% of the scientific papers and patents on non-thermal food processing technologies are directly related with HPP. The jam was the first commercial HPP-processed food introduced into the markets in 1990 followed by retort rice products, cooked hams, and sausages (Yamamoto 2017). HPP is based on the Le Chatelier principle:

"If a dynamic equilibrium is disturbed by changing the conditions, the position of equilibrium moves to counteract the change"

In HPP treatment, isostatic pressure is transmitted to the product through water. Since the water used in HPP is recyclable, it brings about an advantage of reduced energy consumption and environmental protection (Toepfl et al. 2006). The pressure is transmitted uniformly and instantaneously throughout the product, therefore achieving an effect equivalent to pasteurization. A HPP equipment is made up of a high-pressure vessel and its closure, a pressure generator and a material handling mechanism equipped with a temperature control system (Datta & Deeth 1999). The pressure range applied to foods may vary between 300 MPa and 900 MPa depending on the structure and initial microbial load of the product. In practice, the pressure range of 400-600 MPa is sufficient enough to provide microbial safety in many foods (Trujillo 2002). HPP has minimal effect on sensory, nutritional and textural characteristics of the foods (Rendueles et al. 2011; Grundy et al. 2016). The pressure, holding time and temperature applied are the major parameters determining the effectiveness of microbial inactivation.

HPP allows drastically reducing or eliminating the use of preservatives or additives in food. Also, HPP prevents food waste on the retail shelf and in the consumer's refrigerator since it has an extended shelf life (Trujillo et al. 2002). Depending on the processing conditions and product characteristics, the shelf-life of the HPP-treated foods may be extended up to three-fold. HPP may be applied to pre-packed liquid or solid foods. Some high-pressure equipment are suitable for continuous production as well. In the latter case, an aseptic filling may be required depending on the shelf-life expectations and/or safety requirements of the end product. Although HPP has many advantages over traditional heating systems regarding microbial inactivation, it is not effective in the elimination of spores (Pinto et al. 2020). Also, HPP may cause some textural and colour changes in foods under question. Finally, HPP technology is not recommended for the processing of dry products and to ensure microbiological safety, a minimum water activity of 0.8 in foods is required.

Non-thermal food processing technologies are accepted as novel technologies in many countries. Some countries mandate risk analysis of the products manufactured using one or more of the non-thermal food processing technologies. European Union (EU) accepts HPP technology as a 'novel' food processing technology (EC 258/97). As in the standard applications of the EU, the regulations are in the form of 'roof regulation' and the applicability of novel technologies including the implementation details are determined by country regulations. At this point, a position paper published by French authorities in 2010 on HPP technology includes the statement that applications performed at room temperature and around 5 minutes up to 600 MPa pressure are harmless (Jung & Tonello-Samson 2018). This means no risk analysis is required for such foods as long as the conditions stated above are strictly followed. According to the EU framework agreement (European Community Treaty), products produced in any community member country have the right of free circulation within the EU. Today, according to "Regulation of the European Parliament and of the Council on novel foods, amending Regulation (EU) No 1169/2011 of the European Parliament and of the Council and Repealing Regulation (EC) No 258/97 of the European Parliament and of the Council and Commission Regulation (EC) No 1852/2001", application of HPP in food processing is allowed. By now, no chemical and/or microbiological risk that is directly associated with HPP application has been reported. While the number of industrial-scale HPP equipment was 2 in 1994, this figure has reached just over 600 today. In the dairy industry, HPP is being currently used to process cheese milk (Pastoret brand Spanish cheese) or packaged cheese after production (Mu brand Cheddar sticks and Duetto Hot Pepperoni + cheese bars, United Kingdom). In Mexico and Lebanon, two companies are using HPP at the industrial scale to extend the shelf-life of vacuum-packed cheeses. In Australia, cold-pressed milk equivalent to heat-pasteurized milk is being produced commercially (Made by Cow[®], Australia). In New Zealand, a bovine colostrum product branded as Col+[®] is being manufactured by directly processing colostrum with HPP. The last two examples are devoid of any form of heat treatment and approved as safe by country authorities.

2. HHP Treatment of Milk

2.1. Effects of HPP on microbial inactivation in milk

Milk and dairy products are suitable mediums for the growth and survivability of a range of contaminant microorganisms. The degree of contamination and the processing temperature in different milk processing stages significantly determine the diversity of the microbial community in milk (Dash et al. 2022).

Some spoilage and pathogenic microorganisms including Shiga toxin-producing *Escherichia coli, Salmonella* spp., *Listeria monocytogenes, Campylobacter* spp., and *Yersinia* spp. which are known to cause food-borne infections, intoxications and toxicoinfections in humans, are frequently associated with milk and milk products (Dhanashekar et al. 2012; EFSA 2016; Melini et al. 2017; Lee et al. 2019). Therefore, it is important to take every possible measure to avoid such microbiological safety hazards in industrial food processing. The application of HPP generally damages microbial cell walls and membranes, inactivates enzymes,

degrades chromosome DNA, and unfolds and/or dissociates proteins, causing gelation, especially, at higher pressures (i.e., >600 MPa) (Chiozzi et al. 2022; Yang et al. 2012).

It is known that bacterial cells, yeasts and molds are more sensitive to pressure than spores. While pressure treatment at 400-600 MPa at the ambient temperatures inactivate the former microbial groups, much higher pressures and/or longer treatment time are required for the elimination of spores. Specifically, pressure conditions higher than 1200 MPa are considered capable of inactivating spores with relative success; but this very high pressure is not preferred by the food industry since gelation or textural weaknesses in the products are likely to occur. In general, Gram-positive bacteria (*i.e., Listeria monocytogenes, Staphylococcus aureus*) are more resistant to high pressure than Gram-negative bacteria (*i.e., Pseudomonas, Salmonella* spp., *Yersinia enterocolitica, Vibrio parahaemolyticus*). This is due to the presence of teichoic acid in the peptidoglycan structure of gram-positive cell wall, giving cell wall a more rigid structure (Katsaros et al. 2016).

The degree of resistance of milk associated pathogenic and/or spoilage microorganisms against high pressure is affected by multiple factors such as species and strains, bacterial shape, inoculum level, physiological state of bacteria, milk composition, processing pressure, holding time, temperature, and growth phase (Zagorska et al. 2021; Serna-Hernandez et al. 2021).

HPP treatment of donkey milk at 400 MPa for 180 s resulted in <10 colony forming unit/mL of *Pseudomonas* spp., *Enterobacteriaceae*, and *Bacillus cereus* after 30-day storage at 4 °C (Giacometti et al. 2016). HPP application to goat's milk at 600 MPa for 7 minutes at 15 °C did not cause an increase in coliforms, *B. cereus*, yeast and molds during 22 days of storage at 8 °C (Tan et al. 2020). HPP treatment of milk at 345 MPa for 5 min at 50 °C was reported to result in 8 log reduction of *E. coli* and *L. monocytogenes*; however, the reduction in *S. aureus* counts was rather limited (5.33 log) under the same conditions (Alpas et al. 2000).

A summary of the recent studies on the pressure resistance of microorganisms commonly found in different milk categories is given in Table 1. On the other hand, potential impacts of HPP treatment on microorganisms that are less frequently associated with milk such as *Staphylococcus aureus*, *Coxiella burnetii*, and *Mycobacterium*, have been subjected to limited number of scientific evaluations. In a recent project run by Özer et al. (2022), the effectiveness of HPP treatment at 200, 400 or 600 MPa for 5 or 10 min on inactivation of *Mycobacterium tuberculosis*, *Eschericia coli* O157:H7 and spore-forming *Bacillus* spp. in bovine colostrum microfiltrate was investigated. Results demonstrated that except for the treatment at 200 MPa for 5 or 10 min, all other pressure conditions totally inactivated the target pathogens. On the other hand, HPP at 200 MPa for 5 or 10 min failed to inactivate *E. coli* O157:H7 and *M. tuberculosis*. Recently, Yang et al. (2020) showed that multi-cycle HPP treatment (2 x 2.5 min at 600 MPa) resulted in higher level of microbial eradication with satisfactory level of preservation of milk quality than single-cycle treatment (1 x 5 min at 600 MPa). Extra care must be taken in the eradication of pathogenic microorganisms by HPP as their toxins may withstand HPP conditions applied. For example, the number of *Aeromonas hydrophila* AH191 may well be reduced by at least 9 orders of magnitude after HPP treatment at 250 MPa for 30 min at 25 °C, but their toxins may remain unaltered, as concluded by Durães-Carvalho et al. (2012).

2.2. Effect of HPP on milk compounds

Due to the high water activity (i.e., >0.9 a_w), most dairy products including raw or processed milk and fermented dairy products are classified as highly vulnerable foods with a short shelf-life. The HPP successfully extend the shelf-life of milk and milk products without impairing the physical and/or organoleptical characteristics of end product to a large extent (Wang et al. 2016). However, milk components, including fat, casein, whey proteins, enzymes, and minerals can be affected by HHP treatments (Ravash et al. 2022). The method and conditions of processing or the type of milk affect the changes in milk fat. Huppertz et al. (2011) reported that high pressures treatments of cow's milk at 100-600 MPa at 20 °C for 60 min do not influence fat globules. Kiełczewska et al. (2020) demonstrated that HPP treatment of caprine milk at 200-500 MPa for 10 min at 20 °C did not affect the particle size during storage, the colour values, and the overall fatty acids (FAs) profile, but the ratio of branched chain FAs increased. A decrease in FAs upon high pressure treatment to ewe's milk was reported by Gervilla et al. (2001). Milk fat globule membrane size in the range of 1-2 µm tended to increase but those in the range of 2-10 µm decreased after being pressurized at 100-500 MPa at 25 or 50 °C.

HHP may cause modifications in milk proteins. Depending on the HHP conditions, casein micelles may be disintegrated into smaller sub-micelles which are further re-associated (Anema et al. 2005a; Huppertz et al. 2004ab; Orlien 2021; Cadesky et al. 2017). While pressures between 100-200 MPa have limited (if any) modifications in casein micelles, pressures around 250 MPa led to the aggregation of casein micelles and increased the average size of micelles (approximately 25%). Pressures above 400 MPa resulted in breakdown in hydrophobic interactions and decreased the average size of micelles (up to 50%) (Bravo et al. 2015). Whey proteins, especially β -lactoglobulin, may undergo a pressure-driven denaturation (Anema et al. 2005b; Lopez-Fandiño et al. 1996). Whey proteins

Dairy products	Pressure	Exposure time	Temperature	Effect	Reference
Raw milk	600 MPa	5 min	18 °C	5 log reductions for <i>E. coli</i> , <i>Sal-</i> monella and <i>L. monocytogenes</i>	Stratakos et al. (2019)
Cow and goat milks	450 MPa	7 min	15 °C	Increased shelf life (up to 22 days to 8 °C), with no increase in <i>Ba-cillus cereus</i> , mesophilic aerobic spores, coliform, yeast and mold	Tan et al. (2020)
Raw milk	300 MPa	30 min	25 °C	Inactivation in Salmonella spp., E. coli, Shigella, and Staphyloc- cocus aureus	Yang et al. (2012)
Raw whole milk	600 MPa	5 min	40 °C	3 log reductions for total bacterial count and <i>E. coli</i>	Liu et al. (2020)
UHT whole milk	500 MPa	10 min	25 °C	6.20 log reductions for <i>L. mono-</i> cytogenes	Misiou et al. (2018)
Reconstituted milk powder	500-600 MPa	3-5 min	25 °C	Inactivation of <i>Escherichia coli</i> , <i>Pseudomonas fluorescens</i> and <i>Enterobacter aerogenes</i>	Machado et al. (2019)
				Increases in the levels of micro- organism-derived lipopolysac- charides	
Human milk	593.96 MPa	233 s	25 °C	6-7 log reductions for <i>Bacillus</i> cereus, <i>Bacillus aureus</i>	Rocha-Pimienta et al. (2020)

Table 1. Recent studies of applications of high-pressure processing in different milk categories

UHT: Ultra-high temperature treatments

may undergo pressure-driven denaturation of which level is affected by time, temperature and pH of the milk and milk products being pressurized (Huppertz et al. 2004ab; Hinrichs & Rademacher 2004; Arias et al. 2000). Under relatively harsh the processing conditions (i.e., at >300 MPa for >30 min), β -LG irreversibly unfolds which leads to increase in hydrophobicity and hence protein aggregation (Pittia et al. 1996). As the hydrophobic groups buried inside the globular structure of whey proteins become unmasked, the hydrophobicity of the whey proteins increases (Lim et al. 2008). In general, secondary structure of whey proteins is relatively more stable against HPP than the tertiary and quaternary structures (Velez-Ruiz et al. 1998). It has been shown that changes in solubility of whey protein isolate (WPI) is dependent on HPP conditions (Kanno et al. 1998). While no change was observed in solubility of WPI at 400MPa for 10 min, a clear decrease in solubility after HPP at 690 MPa for 5 to 30 min was evident (Lee et al. 2006). Any change in protein leads to a variety of functional characteristics which in turn contribute to the improvement of the organoleptic properties of HPP-treated dairy products (Ravash et al. 2022).

Compared to thermal pasteurization, HPP treatment produces little or no damage on various classes of immunoglobulins (Ig) present in dairy products, which contribute positively to human health (Huang et al. 2020). The HPP treatment of human colostrum at 200 MPa for 2.5, 15 and 30 minutes at 8 °C resulted in insignificant changes in IgA, IgM and IgG (Sousa et al. 2014). It was recently reported that immunoglobulin concentration in human milk remained unchanged after HPP processing at 400 MPa for 5 min and at 593.96 MPa for 233 s (Rocha-Pimienta et al. 2020).

Effects of HPP on milk enzymes show a dependency on HPP conditions. This is an important feature because HPP can be used to control enzymatic activities in dairy products such as mature cheeses, i.e., activation or deactivation of proteolytic and lipolytic enzymes. For example, high pressure application at 400MPa in processing of bovine milk (Munir et al. 2020) and at 200-300 MPa in ewe milk (Alonso et al. 2012) stimulated the proteolysis during cheese maturation (Munir et al. 2020; Alonso et al. 2012).

3. HPP-mediated Chemical and Physicochemical Modifications in Milk and Milk Products

3.1. Effects on milk pH

As a result of increase in the concentration of ionized calcium in milk serum caused by HHP-driven solubilization of micellar calcium phosphate, changes the pH of milk (Huppertz et al. 2004a, 2002; Liepa et al. 2016). The composition of milk-more specifically the buffering capacity of casein micelles- also influences the changes in the pH levels of milk subjected to HPP (Iturmendi et al. 2020). Fat

may have a protective role on HPP-mediated casein micelles' dissociation, thereby diminishing the variation in the milk's pH (Yang et al. 2020; Iturmendi et al. 2020).

3.1.1. Effects on emulsion stability

Emulsion stability is related with the changes in physical parameters of macromolecules including size distribution, flocculation and droplet arrangement over time. Emulsion destabilization is often linked with the changes in physical appearance of a food product, development of undesirable flavors and rapid degradation of nutrients. These changes are due to the exposure of the fat fraction to oxidation and other chemical reactions triggered by emulsion destabilization. HPP affects native milk proteins including ones located on the milk fat globule membrane; hence these modifications affect the emulsion stability of milk. Milder pressure applications can improve emulsion stability *via* exposure to hydrophobic groups *via* the mechanisms explained above. Exposure of hydrophobic groups yields reductions in droplet size as well as modifications in molecular flexibility of milk proteins. This eventually leads to more evenly distribution of polar and non-polar amino acid residues and improves the emulsifying properties. In addition, protein denaturation generates the formation of low-molecular-weight components, and these seem to promote emulsion stability. However, higher pressure treatments, i.e., >400 MPa, seem to adversely affect the emulsification capacity of native milk proteins which leads to decrease in solubility of whey proteins (Gharibzahedi et al. 2019).

3.1.2. Effects on viscosity

While unconcentrated whole and skimmed milk show generally a Newtonian fluid characteristic, concentrated milk display a pseudoplasticity with a shear-thinning flow behavior. Milk viscosity is affected by HPP at varying levels depending on the HPP pressure applied and the time of exposure (Janahar et al. 2021; Serna-Hernandez et al. 2021). Another key factor is the increase in casein micelle hydration caused by HPP-triggered partial disintegration of casein micelles (Zhang et al. 2020).

4. Effect of HPP on Digestion Profiles of Milk Compounds

Although the digestion efficiency of milk components such as proteins and lipid have been well documented (Kopf-Bolanz et al. 2014; Lorieau et al. 2018; Mat et al. 2016; Mulet-Cabero et al. 2019; He et al. 2015), the impact of HPP on digestion profiles of milk components has been subjected to the limited number of scientific studies so far. The effect of HPP on in vitro digestion efficiency of β-lactoglobulin (Maynard et al. 1998; Chicón et al. 2008a), α-casein (Hu et al. 2017), whole milk (Liu et al. 2020) and whey protein isolate (Chicón et al. 2008a) were investigated. The level of tryptic hydrolysis of β -lactoglobulin B increased with increasing pressure and the highest tryptic hydrolysis was obtained at 300 MPa treatment (Stapelfeldt et al. 1996). The peptide profiles of β-lactoglobulin and whey protein isolate treated with HPP in the range of 100-800 MPa and at 400 MPa, respectively, were not affected by the pressure treatment (Maynard et al. 1998; Chicón et al. 2008a). HPP treatment at 200 MPa for 5 min resulted in the highest pepsin digestibility of α -case (Hu et al. 2017) and gastric digestion profiles of whole milk remained unchanged after 600 MPa for 5 min (Liu et al. 2020). As the duration of HPP extends, the digestion efficiency of α -casein decreases (Hu et al. 2017). More recently, Aalaei et al. (2021) investigated the in vitro static gastric digestion profiles of milk proteins subjected to HPP at 400 MPa for 15 min, 600 MPa for 5 or 15 min. The authors used two different static digestion models simulating adult and elderly people's gastric system. Overall, digestion of proteins subjected to HHP at 600 MPa for 5 or 15 min was slower in the elderly model than the adult model evidenced by the high concentration of long chain peptides in the former. Interestingly, HPP at 400 MPa for 15 min improved the protein hydrolysis in the elderly model and yielded more or less similar peptide profiles to the adult model. Increasing pressurizing time at 600 MPa did not cause any further increase in the digestion efficiency of proteins. The majority of the peptides yielded upon in vitro simulated gastric digestion of whey proteins had a length of 16-20 amino acids, indicating high digestion efficiency in whey proteins. In general, caseins, α-lactalbumin and bovine serum albumin are resistant to HHP at 400-600 MPa (Liu et al. 2020; Yang et al. 2020; Lopez-Fandiño et al. 1996). Above 600 MPa, an interaction between caseins and β -lactoglobulin occurs via thiol-disulphide bonds (Bogahawaththa et al. 2018). This eventually slows down the digestion of milk proteins. Regarding milk protein digestion efficiency, 400 MPa seems to be suitable at which microbiological safety is also ensured (Özer et al. 2022). Digestion efficiency of α -casein subjected to HHP at 200 MPa for 5 min decreased by 36-43 % when pressure conditions were set to 600 MPa for 15 min due to casein aggregation (Hu et al. 2017). Similarly, digestion of κ -casein was reduced by half after being processed at 600 MPa (17%) compared with 400 MPa treatment (36%), due possibly to its aggregation with β -lactoglobulin at 600 MPa (Bogahawaththa et al. 2018). In general, although major whey proteins are denatured by HPP, their digestibility does not change at a significant level. Vilela et al. (2006) found that digestibility of single-cycle or triple-cycle pressure-treated WPI with pepsin was higher than untreated WPI after 30 min of digestion (51% and 68% vs 30.9% reduction in WPI, respectively). These findings were further supported by Iskandar et al. (2015) who showed that after 30

min of pancreatic digestion, the degree of pressure-treated WPI hydrolysis reached 95% but the degree of hydrolysis of untreated native WPI was 83%. Most recently, Zhang et al. (2022) demonstrated that the digestion profile and level of retention of nutrients of donor human milk were similar to raw milk but not the samples treated with holder pasteurization at 62.5 °C for 30 min. Protection of lactoferrin in donor human milk by HPP was also higher than those thermally treated (Pitino et al. 2019; Sergius-Ronot et al. 2022). In contrast, a decrease in lactoferrin at very high-pressure applications (550 to 800 MPa) was evident in bovine milk (Bravo et al. 2015). Upon HPP treatment to human donor milk at 350 MPa at 38 °C, the levels of metabolic hormones including insulin, nesfatin-1, cortisol and leptin remained unchanged, glucagon-like peptide 1 level increased and apelin and adiponectin levels decreased (Marousez et al. 2022). However, classical holder pasteurization (62 °C for 30 min) caused dramatic decreases in those metabolic hormones except for adiponectin which remained unchanged.

HPP also effectively reduces the allergenicity of milk proteins depending on the pressurizing conditions (Beran et al. 2009; Huang et al. 2014). In an extensive study, Kleber et al. (2007) demonstrated that the allergenicity of β -lactoglobulin- one of the major allergenic proteins in milk- increased as the pressure and treatment time extended from 200 MPa to 600 MPa and from 0 min to 30 min at <25 °C, respectively. However, when the treatment temperature was increased above 25 °C, the allergenicity of β -lactoglobulin decreased but still higher than the untreated samples. This may be due to the pressure-driven unfolding of whey proteins resulting in the generation of new epitopes which are buried in three-dimensional (3D) structure of folded native proteins (Mills & Mackie 2008). On the other hand, there is no clear correlation between the degree of protein denaturation and the allergenicity, indicating the complexity of food ingredients in food matrices. In most cases, the combination of HPP and enzymatic treatment (i.e., trypsin or chymotrypsin) is applied to reduce allergenicity of proteins (Chicón et al. 2008b).

5. HPP Treatment of Dairy Products

5.1. Impact of HPP on physical stability of whey-based beverage formulations

Fermented whey-based beverages, a promising way to valorize by-products of dairy manufacturing, are often associated with limited shelf-life due to the post-acidification occurring during storage. Time-dependent sedimentation as a result of heat-induced denaturation of whey proteins is another major challenge for whey-based fermented and non-fermented beverages. Severe thermal treatments applied above 70 °C result in protein denaturation, accompanied by loss of aqueous solubility and foaming properties (Kester & Richardson 1984; Pittia et al. 1996). Pega et al. (2018) investigated the effects of HPP at 200 MPa for 10 min or 400 MPa for 1 min on the properties of a fermented beverage manufactured from sweet whey using the starter lactic acid bacteria *Streptococcus thermophilus* and *Lactobacillus delbrueckii* subsp. *bulgaricus*. The authors concluded that the flavor and texture of beverages treated by HPP were maintained up to 45 days post-processing with no changes in chromatic parameters. Sampedro et al. (2009) studied the effects of heat treatment, PEF and HPP processing on pectin methyl esterase (PME) activity and levels of volatile compounds in an orange juice - milk mix beverage. The conditions for inactivation of PME at >90% were as follows: thermal treatment at 85 °C for 1 min, PEF treatment at 25 kV/cm at 65 °C or HPP treatment at 650 MPa at 50 °C. After HPP treatment, the average losses of volatile compounds were between 14.2% and 7.5% at 30 °C, 22.9%, and 42.3% at 50 °C.

Barba et al. (2012) developed an orange juice-milk beverage using HPP at 100-400 MPa for 2 to 9 min. The authors demonstrated that the loss of ascorbic acid in the beverages was <10%, soon after HPP treatment and high-pressure treatment time had no significant effect on ascorbic acid losses. Similar results were reported by Bull et al. (2004) who showed that the ascorbic acid concentration of fruit and vegetable juices was not influenced significantly by HPP at mild temperatures. On the contrary, the colour changes in the HPP-treated samples were more remarkable as the pressure and treatment time increased (Barba et al. 2012).

HPP was demonstrated to better protect the antioxidant capacity of whey-based sweet lime beverages compared with thermal treatment (45.8% vs 76.7%). Sensory parameters of the same beverage remained unchanged during storage as well (Bansal et al. 2019).

5.2. Cheese

HPP has been demonstrated to affect the enzymatic coagulation time, acceleration of ripening, increasing of yield, and modifications to physicochemical and sensory properties of cheese (Chawla et al. 2011; San Martín-González et al. 2006; Naik et al. 2013; López-Pedemonte et al. 2007; Huppertz et al. 2002; Lopez-Fandiño et al. 1996; Nuñez et al. 2020; Chopde et al. 2014; Martínez-Rodríguez et al. 2012; Costabel et al. 2016). Early studies showed that HPP treatment to milk resulted in increased yield in Cheddar (Drake et al. 1997) and semi-hard goat cheese (Trujillo et al. 1999) without impairing the cheese flavor compared to those made from pasteurized milk. There is no clear consensus on the effects of HPP on cheese quality and processing time. For example, Delgado

et al. (2011) demonstrated that the original cheese flavor was not maintained when HPP was applied to cheese made from raw milk at the early stages of maturation. However, this does not mean the development of inferior quality in the end product. Escobedo-Avellaneda et al. (2021) failed to detect any impact of HPP treatment on the coagulation time of milk in Oaxaca cheese production (a pasta-filata type variety). As discussed above, HPP triggers interactions between β -lactoglobulin and κ -casein, leading to retarding caseinomacropeptide release by chymosin. Milder pressure treatments, i.e., <200 MPa cause a reduction in rennet coagulation time of milk. This situation is possibly associated with the increase in the surface area of the casein micelles due to the decrease in the size of the case in micelle with the effect of high pressure, and thus the expansion of the area for the action of chymosin (San Martín-González et al. 2006; Naik et al. 2013). Accelerating cheese ripening without impairing the quality characteristics of cheese is highly desirable by cheesemakers. HPP has been proved to have positive effects on accelerating cheese ripening without altering the quality and sensorial attributes (Chopde et al. 2014; San Martín-González et al. 2006). Cheese-ripening involves a series of complex biochemical reactions mainly regulated by milk enzymes, and proteolysis is the most important biochemical event largely determining the flavor and texture changes in cheese (San Martín-González et al. 2006). The high pressure alters the bacterial cell wall as discussed earlier and modifies the casein matrix, making it more susceptible to proteolytic enzymes activities. Also, HPP-triggered bacterial lysis results in release of bacterial enzymes at higher rates. In addition, the shifts in pH levels and modification of water distribution provide better conditions for enzymatic activity (Chopde et al. 2014; Martínez-Rodríguez et al. 2012). Acceleration of goat's milk cheese ripening by HPP at 400 MPa was also reported by Saldo et al (2000), but the HPP-treated cheeses had crumbly body and bitter flavor defects compared to the untreated cheeses.

The HPP-treated cheeses when compared to traditionally processed cheeses have smaller and uneven compartments in their matrices; however, during the aging process, the differences are almost non-existent, and the final products are relatively similar (Nuñez et al. 2020). Additionally, the interior color of the cheeses is modified by HPP treatment. HPP treatment causes a loss of brightness and an increase in yellowness in the cheeses. Readers are recommended to refer to Nuñez et al. (2020) for an extensive review on the effects of HPP on cheese characteristics.

5.3. Yogurt & fermented milks

The effect of HPP treatment on acid-type dairy gels has been subjected in many scientific reports up to now (Ferragut et al. 2000; Harte et al. 2002; Lanciotti et al. 2004; Vieira et al. 2019; Walsh-O'Grady et al. 2001). In general, within the pressure range of 300-700 MPa, the rheological properties of yogurt-type gels improve which eliminates the necessity for stabilizers and contributes to clean label productions (Loveday et al. 2013). Formation of yogurt matrix relies mainly on the interactions between denatured whey proteins and caseins *via* thiol-disulfide bonds. During the fermentation, the pH is reduced by the action of yogurt starter bacteria to the isoelectric point of caseins and a 3D gel matrix consisting of interacted milk proteins is formed (Soukoulis et al. 2007; Harte et al. 2003). The viscosity of yogurt samples made from milk treated with high pressure treatment at 676 MPa for 30 min had similar rheological and water holding properties to gels made from the reaggregation of disrupted micellar fragments by high pressure during fermentation process. On the other hand, when skim milk was supplemented with whey protein hydrolysate or concentrate (whey protein concentrate-80) prior to HPP treatment, the resulting yogurt had inferior gel properties (Sakkas et al. 2019).

HPP reduces syneresis, a common defect in yogurt and causes thicker and smoother body in yogurt. In addition, in yogurt treated with HPP, the color characteristics -specifically to the L* and b* values- are altered, and the yield stress and the water holding capacity of yogurt matrix are increased. Regarding lipolysis and proteolysis, no clear differences between HPP-treated and thermally treated yogurt samples were reported (Walker et al. 2006; Harte et al. 2003).

Kefir made from HPP-treated whole milk had lower elastic and viscous characteristics, and lightness and color intensity than untreated control and HPP-treated kefir made from skim milk (Renes et al. 2020).

6. Conclusion

Although the advantageous of HPP of foods to food processors, consumers and environment are beyond doubt, more scientific evidence is required to be confident about its chemical safety. Majority of the studies, so far, have focused on the microbiological safety of the food processed by HPP. However, chemical interactions triggered by HPP are yet to be evaluated more deeply. In many countries, food regulations directly related with non-thermal food processing technologies are lacking. This eventually limits the level

of industrialization of HPP and similar non-thermal food processing technologies. Also, the effects of HPP on digestion efficiency of milk compounds deserve more attention.

References

- Aalaei K, Khakimov B, de Gobba C & Ahrne L (2021). Gastric digestion of milk proteins in adult and elderly: effect of high-pressure processing. Foods 10(4): 786. doi.org/10.3390/foods10040786
- Alexandre E M C, Brandão T R S & Silva C L M (2012). Efficacy of non-thermal technologies and sanitizer solutions on microbial load reduction and quality retention of strawberries. *Journal of Food Engineering* 108(3): 417-426. doi.org/10.1016/j.jfoodeng.2011.09.002
- Alonso R, Picon A, Gaya P, Fernández-García E & Nuñez M (2012). Effect of high-pressure treatment of ewe raw milk curd at 200 and 300 MPa on characteristics of Hispanico cheese. *Journal of Dairy Science* 95(7): 3501-3513. doi.org/10.3168/jds.2011-4979
- Alpas H, Kalchayanand N, Bozoglu F & Ray B (2000). Interactions of high hydrostatic pressure, pressurization temperature and pH on death and injury of pressure-resistant and pressure-sensitive strains of foodborne pathogens. *International Journal of Food Microbiology* 60(1): 33-42. doi. org/10.1016/s0168-1605(00)00324-x
- Anema S G, Lowe E K & Stockmann R (2005a). Particle size changes and casein solubilisation in high-pressure-treated skim milk. Food Hydrocolloids 19: 257-267. doi.org/10.1016/j.foodhyd.2004.04.025
- Anema S G, Stockman R & Lowe E K (2005b). Denaturation of beta-lactoglobulin in pressure-treated skim milk. *J Agric Food Chem* 53(20): 7783-7791. doi.org/10.1021/jf050326x
- Arias R, Lee T-C, Logendra L & Janes H (2000). Correlation of lycopene measured by HPLC with the L*, a*, b* color readings of a hydroponic tomato and the relationship of maturity with color and lycopene content. J Agric Food Chem 48(5): 1697-1702. doi.org/10.1021/jf990974e
- Bansal V, Jabeen K, Rao P S, Prasad P & Yadav K (2019). Effect of high pressure processing (HPP) on microbial safety, physicochemical properties, and bioactive compounds of whey-based sweet lime (whey-lime) beverage. *Journal of Food Measurement and Characterization* 13(4): 454-465. doi.org/10.1007/s11694-018-9959-1
- Barba F J, Cortés C, Esteve M J & Frígola A (2012). Study of antioxidant capacity and quality parameters in an orange juice-milk beverage after highpressure processing treatment. Food Bioprocess Technology 5: 2222-2232. doi.org/10.1007/s11947-011-0570-2
- Beran M, Klubal R, Molik P, Strohalm J, Urban M, Klaudyova A A & Prajlerova K (2009). Influence of high-hydrostatic pressure on tryptic and chymotryptic hydrolysis of milk proteins. *High Pressure Research* 29(1): 23-27. doi.org/10.1080/08957950802492142
- Bogahawaththa D, Buckow R, Chandrapala J & Vasiljevic T (2018). Comparison between thermal pasteurization and high pressure processing of bovine skim milk in relation to denaturation and immunogenicity of native milk proteins. *Innovative Food Science and Emerging Technologies* 47: 301-308. doi.org/10.1016/j.ifset.2018.03.016
- Bravo F I, Felipe X, López-Fandino R & Molina E (2015). Skim milk protein distribution as a result of very high hydrostatic pressure. *Food Research International* 72: 74-79. doi.org/10.1016/j.foodres.2015.03.014
- Bull M K, Zerdin K, Howe E, Goicoechea D, Paramanandhan P, Stockman R, Sellahewa J, Szabo E A, Johnson R L & Stewart C M (2004). The effect of high pressure processing on the microbial, physical and chemical properties of Valencia and Navel orange juice. *Innovative Food Science and Emerging Technologies* 5(2): 135-149. doi.org/10.1016/j.ifset.2003.11.005
- Cadesky L, Walkling-Ribeiro M, Kriner K T, Karwe M V & Moraru C I (2017). Structural changes induced by high-pressure processing in micellar casein and milk protein concentrates. *Journal of Dairy Science* 100(9): 7055-7070. doi.org/10.3168/jds.2016-12072
- Chawla R, Patil G R & Singh A K (2011). High hydrostatic pressure technology in dairy processing: A review. *Journal of Food Science and Technology* 48: 260-268. doi.org/10.1007/s13197-010-0180-4
- Chicón R, Belloque J, Alonso E & López-Fandiño R (2008a). Immunoreactivity and digestibility of high-pressure-treated whey proteins. *International Dairy Journal* 18(4): 367-376. doi.org/10.1016/j.idairyj.2007.11.010
- Chicón R, Belloque J, Alonso E, Martín-Alverez P J & López-Fandiño R (2008b). Hydrolysis under high hydrostatic pressure as a means to reduce the binding of β-lactoglobulin to immunoglobulin E from human sera. J Food Prot 71(7): 1453-1459. doi.org/10.4315/0362-028x-71.7.1453
- Chiozzi V, Agriopoulou S & Varzakas T (2022). Advances, applications, and comparison of thermal (pasteurization, sterilization, and aseptic packaging) against non-thermal (ultrasounds, UV radiation, ozonation, high hydrostatic pressure) technologies in food processing. *Appl Sci*, 12(4): 2202. doi.org/10.3390/app12042202
- Chopde S S, Deshmukh M A, Kalyankar S D & Changade S P (2014). High pressure technology for cheese processing-a review. Asian Journal of Dairy & Food Research 33(4): 239-245. doi.org/10.5958/0976-0563.2014.00610.1
- Costabel L M, Bergamini C, Vaudagna S R, Cuatrin A L, Audero G & Hynes E (2016). Effect of high-pressure treatment on hard cheese proteolysis. *Journal of Dairy Science* 99(6): 4220-4232. doi.org/10.3168/jds.2015-9907
- Dash K K, Fayaz U, Dar A H, Shams R, Manzoor S, Sundarsingh A, Deka P & Khan S A (2022). A comprehensive review on heat treatments and related impact on the quality and microbial safety of milk and milk-based products. *Food Chemistry Advances* 1: 100041. doi.org/10.1016/j. focha.2022.100041
- Datta N & Deeth H C (1999). High pressure processing of milk and dairy products. Australian Journal of Dairy Technology 54(1): 41-48.
- Delgado J F, González-Crespo J, Cava R & Ramírez R (2011). Changes in the volatile profile of a raw goat milk cheese treated by hydrostatic high pressure at different stages of maturation. *International Dairy Journal* 21(3): 135-141. doi.org/10.1016/j.idairyj.2010.10.006

- Dhanashekar R, Akkinepalli S & Nellutla A (2012). Milk-borne infections. An analysis of their potential effect on the milk industry. *Germs* 2(3): 101-109. doi.org/10.11599/germs.2012.1020
- Drake M A, Harrinson S L, Asplund M, Barbosa-Canovas G & Swanson B G (1997). High pressure treatment of milk and effects on microbiological and sensory quality of Cheddar cheese. *Journal of Food Science* 62(4): 843-860. doi.org/10.1111/j.1365-2621.1997.tb15468.x
- Durães-Carvalho R, Souza A R, Martins M L, Sprogis A C S, Bispo J A C, Bonafe C F S & Yano T (2012). Effect of high hydrostatic pressure on *Aeromonas hydrophila* AH 191 growth in milk. *Journal of Food Science* 77(8): M417-M424. doi.org/10.1111/j.1750-3841.2012.02819.x
- EFSA (European Food Safety Authority) (2016). The European Union summary report on trends and sources of zoonoses, zoonotic agents and foodborne outbreaks in 2015. EFSA Journal 14(12): 4634. doi.org/10.2903/j.efsa.2016.4634
- Escobedo-Avellaneda Z, Espricueta-Candelaria R S, Calvo-Segura S, Welti-Chanes J & Chuck-Hernández C (2021). Changes induced by high hydrostatic pressure in acidified and non-acidified milk during Oaxaca cheese production. *International Journal of Food Science & Technology* 56(9): 4639-4649. doi.org/10.1111/jjfs.15134
- Ferragut V, Martínez V M, Trujillo A J & Guamis B (2000). Properties of yogurts made from whole ewe's milk treated by high hydrostatic pressure. *Milchwissenchaft* 55(5): 267-269.
- Gervilla R, Ferragut V & Guamis B (2001). High hydrostatic pressure effects on color and milk fat globule of ewe's milk. *Journal of Food Science* 66(6): 880-885. doi.org/10.1111/j.1365-2621.2001.tb15190.x
- Giacometti F, Bardasi L, Merialdi G, Morbarigazzi M, Federici S, Piva S & Serraino A (2016). Shelf life of donkey milk subjected to different treatment and storage conditions. *Journal of Dairy Science* 99(6): 4291-4299. doi.org/10.3168/jds.2015-10741
- Gharibzahedi S M T, Hernández-Ortega C, Welti-Chanes J, Putnik P, Barba F J, Mallikarjunan K, Escobedo-Avellaneda Z & Roohinejad S (2019). High pressure processing of food-grade emulsion systems: Antimicrobial activity, and effect on the physicochemical properties. *Food Hydrocolloids* 87: 307-320. doi.org/10.1016/j.foodhyd.2018.08.012
- Grundy M M L, Lapsley K & Ellis P R (2016). A review of the impact of processing on nutrient bioaccessibility and digestion of almonds. *International Journal of Food Science & Technology* 51(9): 1937-1946. doi.org/10.1111/ijfs.13192
- Harte F M, Amonte L, Luedecke B G, Swanson B G & Barbosa-Cánovas G V (2002). Yield stress and microstructure of set yogurt made from high hydrostatic pressure-treated full fat milk. *Journal of Food Science* 67(6): 2245-2250. doi.org/10.1111/j.1365-2621.2002.tb09535.x
- Harte F, Luedecke L, Swanson B & Barbosa-Cánovas G V (2003). Low-fat set yogurt made from milk subjected to combinations of high hydrostatic pressure and thermal processing. *Journal of Dairy Science* 86(4): 1074-1082. https://doi.org/10.3168/jds.s0022-0302(03)73690-x
- He Z, Yuan B, Zeng M, Tao G & Chen J (2015). Effect of simulated processing on the antioxidant capacity and in vitro protein digestion of fruit juicemilk beverage model systems. *Food Chemistry* 175: 457-464. doi.org/10.1016/j.foodchem.2014.12.007
- Hinrichs J & Rademacher B (2004). High pressure thermal denaturation kinetics of whey proteins. *Journal of Dairy Research* 71(4), 480-488. doi. org/10.1017/s0022029904000238
- Hu G, Zheng Y, Liu Z, Xiao Y, Deng Y & Zhao Y (2017). Effects of high hydrostatic pressure, ultraviolet light-C, and far-infrared treatments on the digestibility, antioxidant and antihypertensive activity of α-casein. Food Chemistry 221: 1860-1866. doi.org/10.1016/j.foodchem.2016.10.088
- Huang H W, Hsu C P, Yang B B & Wang C Y (2014). Potential utility of high pressure processing to address the risk of food allergen concerns. *Comprehensive Reviews in Food Science and Food Safety* 13: 78-90. https://doi.org/10.1111/1541-4337.12045
- Huang H W, Hsu C P & Wang C Y (2020). Healthy expectations of high hydrostatic pressure treatment in food processing industry. *Journal of Food and Drug Analysis* 28(1): 1-13. doi.org/10.1016/j.jfda.2019.10.002
- Huppertz T, Kelly A L & Fox P F (2002). Effects of high pressure on constituents and properties of milk. *International Dairy Journal* 12(7): 561-572. doi.org/10.1016/s0958-6946(02)00045-6
- Huppertz T, Fox P F & Kelly A L (2004a). High pressure treatment of bovine milk: Effects on casein micelles and whey proteins. *Journal of Dairy Research* 71(1): 97-106. doi.org/10.1017/s002202990300640x
- Huppertz T, Fox P F & Kelly A L (2004b). Properties of casein micelles in high pressure-treated bovine milk. Food Chemistry 87(1): 103-110. doi. org/10.1016/j.foodchem.2003.10.025
- Huppertz T, Smiddy M A, Goff H D & Kelly A L (2011). Effects of high pressure treatment of mix on ice cream manufacture. *International Dairy Journal* 21(9): 718-726. doi.org/10.1016/j.idairyj.2010.12.005
- Iskandar M M, Lands L C, Sabally K, Azadi B, Meehan B, Mawji N, Skinner C D, Kubow S (2015). High hydrostatic pressure pretreatment of whey protein isolates improves their digestibility and antioxidant capacity. *Foods* 4(2): 184-207. doi.org/10.3390/foods4020184
- Iturmendi N, García A, Galarza U, Barba C, Fernández T & Maté J I (2020). Influence of high hydrostatic pressure treatments on the physicochemical, microbiological and rheological properties of reconstituted micellar casein concentrates. *Food Hydrocolloids* 106: 105880. doi.org/10.1016/j. foodhyd.2020.105880
- Janahar J J, Marciniak A, Balasubramaniam V M, Jimenez-Flores R. & Ting E (2021). Effects of pressure, shear, temperature, and their interactions on selected milk quality attributes. *Journal of Dairy Science* 104(2): 1531–1547. doi.org/10.3168/jds.2020-19081
- Jung S & Tonello-Samson C (2018). High hydrostatic pressure food processing: Potential and limitations. In: Proctor A, (Ed.), Alternatives to Conventional Food Processing (2nd ed.), Royal Society of Chemistry, London, pp. 251-315. doi.org/10.1039/9781782626596-00251
- Kanno C, Mu T-H, Hagiwara T, Ametani M & Azuma N (1998). Gel formation from industrial milk whey proteins under hydrostatic pressure: effect of hydrostatic pressure and protein concentration. *Journal of Agricultural and Food Chemistry* 46(2): 417-424. doi.org/10.1021/jf970652f

- Katsaros G, Alexandrakis Z & Taoukis P (2016). High-pressure processing of foods technology and application. In: T Varzakas & C Tzia, (Eds.), *Handbook of Food Processing: Food Preservation*, CRC Press, Boca Raton, FL, pp. 443-468.
- Kester J J & Richardson T (1984). Modification of whey proteins to improve functionality. J Dairy Sci 67: 2757-2774. doi.org/10.3168/jds.s0022-0302(84)81633-1
- Kiełczewska K, Jankowska A, Dąbrowska A, Wachowska M & Ziajka J (2020). The effect of high pressure treatment on the dispersion of fat globules and the fatty acid profile of caprine milk. *International Dairy Journal* 102: 104607. doi.org/10.1016/j.idairyj.2019.104607
- Kleber N, Maier S & Hinrichs J (2007). Antigenic response of bovine β-lactoglobulin influenced by ultra-high pressure treatment and temperature. Innovative Food Science and Emerging Technologies 8(1): 39-45. doi.org/10.1016/j.ifset.2006.05.001
- Kopf-Bolanz K A, Schwander F, Gijs M, Vergères G, Portmann R & Egger L (2014). Impact of milk processing on the generation of peptides during digestion. *International Dairy Journal* 35(2): 130-138. doi.org/10.1016/j.idairyj.2013.10.012
- Lanciotti R, Vannini L, Pittia P & Guerzoni M E (2004). Suitability of high-dynamic-pressure-treated milk for the production of yoghurt. Food Microbiology 21(6): 753-760. doi.org/10.1016/j.fm.2004.01.014
- Lee W, Clark S & Swanson B G (2006). Functional properties of high hydrostatic pressure-treated whey protein. *Journal of Food Processing and Preservation* 30(4): 488-501. doi.org/10.1111/j.1745-4549.2005.00081.x
- Lee S H I, Cappato L P, Guimarães J T, Balthazar C F, Rocha R S, Franco L T, da Cruz A G, Corassin C H & de Oliveira C A F (2019). Listeria monocytogenes in milk: occurrence and recent advances in methods for inactivation. Beverages 5(1): 14. doi.org/10.3390/beverages5010014
- Liepa M, Zagorska J, Galoburda R (2016). High-pressure processing as novel technology in dairy industry: a review. *Research for Rural Development* 1: 76-83.
- Lim S-Y, Swanson B G & Clark S (2008). High hydrostatic pressure modification of whey protein concentrate for improved functional properties. Journal of Dairy Science 91(4): 1299-1307. doi.org/10.3168/jds.2007-0390
- Liu G, Carøe C, Qin Z, Munk D M E, Crafack M, Petersen M A & Ahrné L (2020). Comparative study on quality of whole milk processed by high hydrostatic pressure or thermal pasteurization treatment. *LWT* 127: 109370. doi.org/10.1016/j.lwt.2020.109370
- Lopez-Fandiño R, Carrascosa A V & Olano A (1996). The effects of high pressure on whey protein denaturation and cheese-making properties of raw milk. J Dairy Sci 79: 929-936. doi.org/10.3168/jds.s0022-0302(96)76443-3
- López-Pedemonte T, Roig-Sagués A X, de Lamo S, Gervilla R & Guamis B (2007). High hydrostatic pressure treatment applied to model cheeses made from cow's milk inoculated with *Staphylococcus aureus*. *Food Control* 18: 441-447. doi.org/10.1016/j.foodcont.2005.11.012
- Lorieau L, Halabi A, Ligneul A, Hazart E, Dupont D & Floury J (2018). Impact of the dairy product structure and protein nature on the proteolysis and amino acid bioaccessibility during *in vitro* digestion. *Food Hydrocolloids* 82: 399-411. doi.org/10.1016/j.foodhyd.2018.04.019
- Loveday S M, Sarkar A & Singh H (2013). Innovative yoghurts: Novel processing technologies for improving acid milk gel texture. *Trends in Food Science and Technology* 33(1): 5-20. doi.org/10.1016/j.tifs.2013.06.007
- Machado K I A, Roquetto A R, Moura C S, de Souza-Lopes A, Cristianini M & Amaya-Farfan J (2019). Comparative impact of thermal and high isostatic pressure inactivation of gram-negative microorganisms on the endotoxic potential of reconstituted powder milk. *LWT* 106: 78-82. doi. org/10.1016/j.lwt.2019.02.064
- Mahalik N P & Nambiar A N (2010). Trends in food packaging and manufacturing systems and technology. *Trends in Food Science & Technology* 21(3): 117-128. doi.org/10.1016/j.tifs.2009.12.006
- Marousez L, Tran L, Micours E, de Lamballerie M, Gottrand F, Pierrat V, Eberlé D, Ley D & Lesage J (2022). Metabolic hormones in human breast milk are preserved by high hydrostatic pressure processing but reduced by Holder pasteurization. *Food Chemistry* 377: 131957. doi.org/10.1016/j. foodchem.2021.131957
- Martínez-Rodríguez Y, Acosta-Muñiz C, Olivas G I, Guerrero-Beltrán J, Rodrigo-Aliaga D & Sepúlveda D R (2012). High hydrostatic pressure processing of cheese. *Comprehensive Reviews in Food Science and Food Safety* 11(4): 399-416. doi.org/10.1111/j.1541-4337.2012.00192.x
- Mat D J L, Le Feunteun S, Michon C & Souchon I (2016). *In vitro* digestion of foods using pH-stat and the INFOGEST protocol: Impact of matrix structure on digestion kinetics of macronutrients, proteins and lipids. *Food Research International* 88: 226-233. doi.org/10.1016/j. foodres.2015.12.002
- Maynard F, Weingand A, Hau J & Jost R (1998). Effect of high-pressure treatment on the tryptic hydrolysis of bovine β-lactoglobulin ab. *International Dairy Journal* 8(2): 125-133. doi.org/10.1016/s0958-6946(98)00030-2
- Melini F, Melini V, Luziatelli F & Ruzzi, M (2017). Raw and heat-treated milk: From public health risks to nutritional quality. *Beverages*, 3(4): 54. doi.org/10.3390/beverages3040054
- Mills E N C & Mackie A R (2008). The impact of processing on allergenicity of food. *Current Opinion in Allergy and Clinical Immunology* 8(3): 249-253. doi.org/10.1097/aci.0b013e3282ffb123
- Misiou O, van Nassau T J, Lenz C A & Vogel R F (2018). The preservation of Listeria-critical foods by a combination of endolysin and high hydrostatic pressure. *International Journal of Food Microbiology* 266: 355-362. doi.org/10.1016/j.ijfoodmicro.2017.10.004
- Mulet-Cabero A -I, Mackie A R, Wilde P J, Fenelon M A & Brodkorb A (2019). Structural mechanism and kinetics of in vitro gastric digestion are affected by process-induced changes in bovine milk. *Food Hydrocolloids* 86: 172-183. doi.org/10.1016/j.foodhyd.2018.03.035

- Munir M, Nadeem M, Mahmood-Qureshi M, Gamlath C J, Martin G J O, Hemar Y & Ashokkumar M (2020). Effect of sonication, microwaves and high-pressure processing on ACE-inhibitory activity and antioxidant potential of Cheddar cheese during ripening. Ultrasonics Sonochemistry 67: 105140. doi.org/10.1016/j.ultsonch.2020.105140
- Naik L, Sharma R, Rajput Y S & Manju G (2013). Application of high pressure processing technology for dairy food preservation -future perspective: a review. J Anim Prod Adv 3(8): 232-241. doi.org/10.5455/japa.20120512104313
- Nuñez M, Calzada J & del Olmo A (2020). High pressure processing of cheese: Lights, shadows and prospects. *International Dairy Journal* 100: 104558. doi.org/10.1016/j.idairyj.2019.104558
- Orlien V (2021). Structural changes induced in foods by HPP. In: K Knoerzer & K Muthukumarappan (Eds.), *Innovative Food Processing Technologies*, Elsevier, Oxford, pp. 112-129. doi.org/10.1016/b978-0-08-100596-5.22685-1
- Özer B, Yazihan N, Hocalar M & Hocalar B (2022). Development of colostrum beverage enriched for functional compounds using high pressure and membrane combinations. TUBITAK-TEYDEB Project (7190157). Unpublished project report.
- Pega J, Denoya G I, Castells M L, Sarquis S, Aranibar G F, Vaudagna S R & Nanni M (2018). Effect of high-pressure processing on quality and microbiological properties of a fermented beverage manufactured from sweet whey throughout refrigerated storage. *Food and Bioprocess Technology* 11: 1101-1110. doi.org/10.1007/s11947-018-2078-5
- Pinto C A, Moreira S, Fidalgo L G, Inácio R S, Barba F J & Saraiva J A (2020). Effects of high-pressure processing on fungi spores: Factors affecting spore germination and inactivation and impact on ultrastructure. *Comprehensive Reviews in Food Science and Food Safety* 19(2): 553-573. doi. org/10.1111/1541-4337.12534
- Pitino M A, Unger S, Doyen A, Pouliot Y, Aufreiter S, Stone D, Kiss A & O'Connor D L (2019). High hydrostatic pressure processing better preserves the nutrient and bioactive compounds composition of human donor milk. *The Journal of Nutrition* 149(3): 497-504. doi.org/10.1093/jn/nxy302
- Pittia P, Wilde J P & Clark D C (1996). The foaming properties of native and pressure treated β-casein. *Food Hydrocolloids* 10(3): 335-342. doi. org/10.1016/s0268-005x(96)80010-5
- Ravash N, Peighambardoust S H, Soltanzadeh M, Pateiro M & Lorenzo J M (2022). Impact of high-pressure treatment on casein micelles, whey proteins, fat globules and enzymes activity in dairy products: A review. *Critical Reviews in Food Science and Nutrition* 62(11): 2888-2908. doi.or g/10.1080/10408398.2020.1860899
- Rendueles E, Omer M K, Alvseike O, Alonso-Calleja C, Capita R & Prieto M (2011). Microbiological food safety assessment of high hydrostatic pressure processing: a review. LWT - Food Science and Technology 44(5): 1251-1260. doi.org/10.1016/j.lwt.2010.11.001
- Renes E, Fernández A, López M & Álvarez-Ordoñez A (2020). Effect of high hydrostatic pressure processing of milk on the quality characteristics of kefir. Journal of Food Processing and Preservation 44: e14797. doi.org/10.1111/jfpp.14797
- Rocha-Pimienta J, Martillanes S, Ramírez R, Garcia-Parra J & Delgado-Adamez J (2020). Bacillus cereus spores and Staphylococcus aureus sub. aureus vegetative cells inactivation in human milk by high-pressure processing. Food Control 113: 107212. doi.org/10.1016/j.foodcont.2020.107212
- Sakkas L, Tzevdou M, Zoidou E, Gkotzia E, Karvounis A, Samara A, Taoukis P & Moatsou G (2019). Yoghurt-type gels from skim sheep milk base enriched with whey protein concentrate hydrolysates and processed by heating or high hydrostatic pressure. *Foods* 8: 342. doi.org/10.3390/ foods8080342
- Saldo J, Sendra E & Guamis B (2000). High hydrostatic pressure for acceleratring ripenin gof goat's milk cheese: proteolysis and texture. *Journal of Food Science* 65(4): 636-640. doi.org/10.1111/j.1365-2621.2000.tb16064.x
- Sampedro F, Geveke D J, Fan X & Zhang H Q (2009). Effect of PEF, HHP and thermal treatment on PME inactivation and volatile compounds concentration of an orange juice-milk based beverage. *Innovative Food Science and Emerging Technologies* 10(4): 463-469. doi.org/10.1016/j. ifset.2009.05.006
- San Martín-González M F, Welti-Chanes J & Barbosa-Cánovas G V (2006). Cheese manufacture assisted by high pressure. *Food Reviews International* 22: 275-289. doi.org/10.1080/87559120600695157
- Sergius-Ronot M, Pitino M A, Suwal S, Shama S, Unger S, O'Connor D L, Pouliot Y & Doyen A (2022). Impact of holder, high temperature short time and high hydrostatic pressure pasteurization methods on protein structure and aggregation in a human milk protein concentrate. *Food Chemistry* 374: 131808. doi.org/10.1016/j.foodchem.2021.131808
- Serna-Hernandez S O, Escobedo-Avellaneda Z, García-García R, Rostro-Alanis M D J & Welti-Chanes J (2021). High hydrostatic pressure induced changes in the physicochemical and functional properties of milk and dairy products: a review. Foods 10(8): 1867. doi.org/10.3390/foods10081867
- Soukoulis C, Panagiotidis P, Koureli R & Tzia C (2007). Industrial yogurt manufacture: Monitoring of fermentation process and improvement of final product quality. *Journal of Dairy Science* 90: 2641-2654. doi.org/10.3168/jds.2006-802
- Sousa S G, Delgadillo I & Saraiva J A (2014). Effect of thermal pasteurisation and high-pressure processing on immunoglobulin content and lysozyme and lactoperoxidase activity in human colostrum. Food Chemistry 151: 79-85. doi.org/10.1016/j.foodchem.2013.11.024
- Stapelfeldt H, Petersen P H, Kristiansen K R, Qvist K B, Skibsted L H (1996). Effect of high hydrostatic pressure on the enzymic hydrolysis of betalactoglobulin B by trypsin, thermolysin and pepsin. *Journal of Dairy Research* 63(1): 111-118. doi.org/10.1017/s0022029900031587
- Stratakos A C, Inguglia E S, Linton M, Tollerton J, Murphy L, Corcionivoschi N, Koidis A & Tiwari B K (2019). Effect of high pressure processing on the safety, shelf life and quality of raw milk. *Innovative Food Science and Emerging Technologies* 52: 325-333. doi.org/10.1016/j.ifset.2019.01.009
- Tan S.F, Chin N L, Tee T P & Chooi S K (2020). Physico-chemical changes, microbiological properties, and storage shelf life of cow and goat milk from industrial high-pressure processing. *Processes* 8(6): 697. doi.org/10.3390/pr8060697

- Toepfl S, Mathys A, Heinz V & Knorr D (2006). Potential of high hydrostatic pressure and pulsed electric fields for energy efficient and environmentally friendly food processing. *Food Reviews International* 22(4): 405-423. doi.org/10.1080/87559120600865164
- Trujillo A J, Royo C, Ferragut V & Guamis B (1999). Ripening profiles of goat cheese produced from milk treated with high pressure. Journal of Food Science 64(5): 833-837. doi.org/10.1111/j.1365-2621.1999.tb15922.x
- Trujillo A J (2002). Applications of high-hydrostatic pressure on milk and dairy products. *High Pressure Research* 22: 619-626. doi. org/10.1080/08957950212449
- Velez-Ruiz J F, Swanson B G & Barbosa-Canovas G (1998). Flow and viscoelastic properties of concentrated milk treated by high hydrostatic pressure. LWT-Food Science and Technology 31(2): 182-195. doi.org/10.1006/fstl.1997.9999
- Vieira P, Pinto C A, Lopes-da-Silva J A, Remize F, Barba F J, Marszałek K, Delgadillo I & Saraiva J A (2019). A microbiological, physicochemical, and texture study during storage of yoghurt produced under isostatic pressure. LWT 110: 152-157. doi.org/10.1016/j.lwt.2019.04.066
- Vilela R M, Lands L C, Chan H M, Azadi B, Kubow S (2006). High hydrostatic pressure enhances whey protein digestibility to generate whey peptides that improve glutathione status in CFTR-deficient lung epithelial cells. *Molecular Nutrition and Food Research* 50(11):1013-1129. doi. org/10.1002/mnfr.200600074
- Walker M K, Farkas D F, Loveridge V, Meunier-Goddik L (2006). Fruit yogurt processed with high pressure. International Journal of Food Science and Technology 41(4): 464-467. doi.org/10.1111/j.1365-2621.2005.01084.x
- Walsh-O'Grady C D, O'Kennedy B T, Fitzgerald R J & Lane C N (2001). A rheological study of acid-set "simulated yogurt milk" gels prepared from heat- or pressure-treated milk proteins. *Lait* 81: 637-650. doi.org/10.1051/lait:2001103
- Wang C Y, Huang H W, Hsu C P, Yang B B (2016). Recent advances in food processing using high hydrostatic pressure technology. Critical Reviews in Food Science and Nutrition 56: 527-540. doi.org/10.1080/10408398.2012.745479
- Yamamoto K (2017). Food processing by high hydrostatic pressure. *Bioscience, Biotechnology, Biochemistry* 81(4): 672-679. doi.org/10.1080/0916 8451.2017.1281723
- Yang B, Shi Y, Xia X, Xi M, Wang X, Ji B & Meng J (2012). Inactivation of foodborne pathogens in raw milk using high hydrostatic pressure. Food Control 28(2): 273-278. doi.org/10.1016/j.foodcont.2012.04.030
- Yang S, Liu G, Munk D M E, Qin Z, Petersen M A, Cardoso D R & Otte J, Ahrné L (2020). Cycled high hydrostatic pressure processing of whole and skimmed milk: Effects on physicochemical properties. *Innovative Food Science and Emerging Technologies* 63: 102378. doi.org/10.1016/j. ifset.2020.102378
- Zagorska J, Galoburda R, Raita S & Liepa M (2021). Inactivation and recovery of bacterial strains, individually and mixed, in milk after high pressure processing. *International Dairy Journal* 123: 105147. doi.org/10.1016/j.idairyj.2021.105147
- Zhang D, Palmer J, Teh K H & Flint S (2020). Identification and selection of heat-stable protease and lipase-producing psychrotrophic bacteria from fresh and chilled raw milk during up to five days storage. *LWT* 134: 110165. doi.org/10.1016/j.lwt.2020.110165
- Zhang J, Le N A, Duley J A, Cowley D M, Shaw P N & Bansal N (2022). Comparing the effects of hydrostatic high-pressure processing vs holder pasteurisation on the microbial, biochemical and digestion properties of donor human milk. *Food Chemistry* 373: 131545. doi.org/10.1016/j. foodchem.2021.131545



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